

ECEC21

Session 2B: Detection, Fate, and Transport of PFAS 2

April 27, 2021 4:30



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Microbiological Signature of PFAS-laden Surface Water Foams

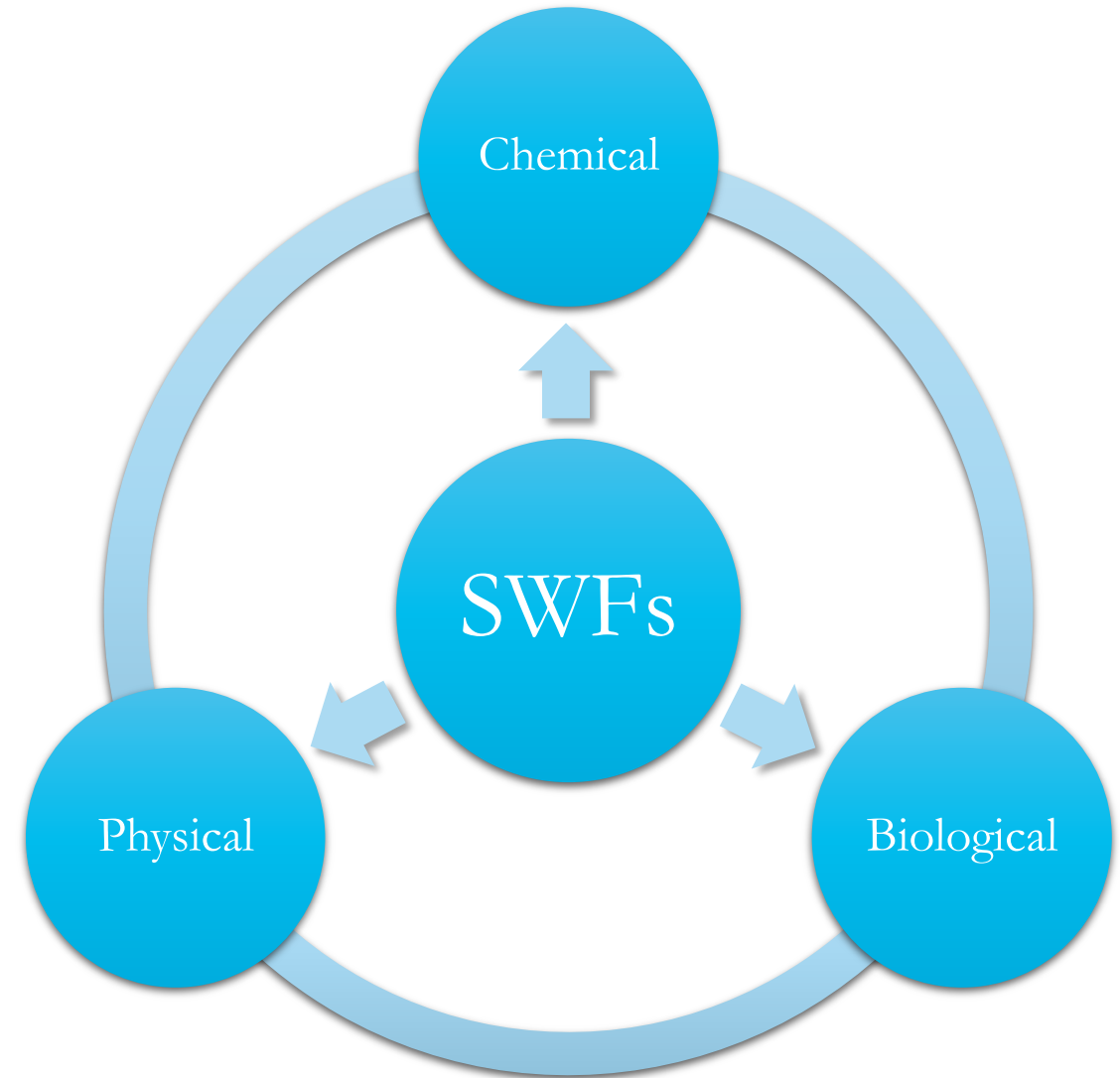
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MICHIGAN DEPARTMENT OF
ENVIRONMENT, GREAT LAKES, AND ENERGY

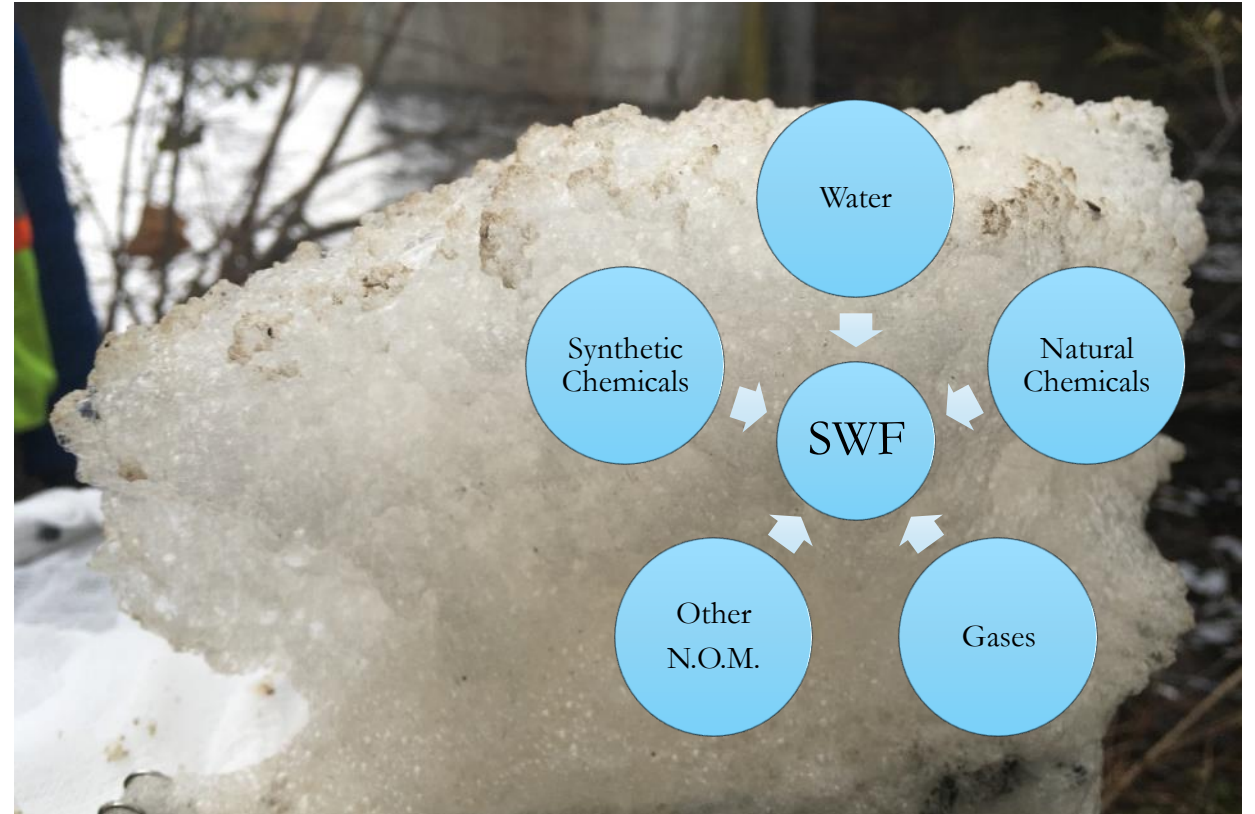
Context

- Michigan Department of Environment, Great Lakes, and Energy (EGLE) & Michigan PFAS Action Response Team (MPART).
- Field Work 2019-2021: Four rivers & three inland lakes.
- First study using extended 41-list of PFAS and Metagenomic Sequencing of SWF.
- AECOM *Surface Water Foam Study Report*, Michigan EGLE, January 7, 2021.



Surface Water Foams (SWFs)

- Can be natural or of human origin or both,
- Globally widespread and form in marine, brackish and freshwater habitats,
- Compositionally are made of water, gases, and mineral fractions, with traces of natural and synthetic chemicals and biological components,
- Genesis, occurrence, fate and transport are on-going study topics by many researchers.



Stable SWF collected from the Rogue River, Michigan [11/04/19].

Precedence: Biodegradation of PFAS

Organism	Compounds	Process	Reference
<i>Pseudomonas parafulva</i>	PFOS, PFOA	Degradation	Yi, et al., 2018
<i>Pseudomonas aeruginosa</i>	PFOS	Degradation	Kwon, et al., 2019
<i>Pseudomonas plecoglossicida</i>	PFOS	Degradation	Chetverikov, et al., 2017
<i>Acidimicrobium</i> sp. strain A6	PFOS, PFOA	Degradation/Defluorination	Huang & Jaffé, 2019
<i>Pseudomonas [striata]</i> strain PS27 <i>[Burkholderia cenocepacia]</i> strain PS27]	PFHxS	Bioaccumulation	Presentato, et al., 2020
<i>Pseudomonas</i> sp. strain PDMF10	PFHxS	Bioaccumulation	Presentato, et al., 2020
<i>Gordonia</i> sp. strain NB4-1Y	6:2 FTSA	Degradation	Shaw, et al., 2019
<i>Desulfococcus</i>	Total PFAS	Positive Correlation	O'Carroll, et al., 2020
<i>Oxalobacteraceae</i>	Total PFAS	Negative Correlation	O'Carroll, et al., 2020
<i>Syntrophaceae</i>	Total PFAS	Positive Correlation	O'Carroll, et al., 2020
<i>Gordonia</i> sp., <i>Nitrosomas</i> , <i>Rhodococcus</i>	4:2, 6:2, 8:2 FTSA	Transformation	Harding-Marjanovic, et al., 2015
Various Fungi [Laccase]	PFOA	Laccase Catalysis Degradation	Luo, et al., 2015

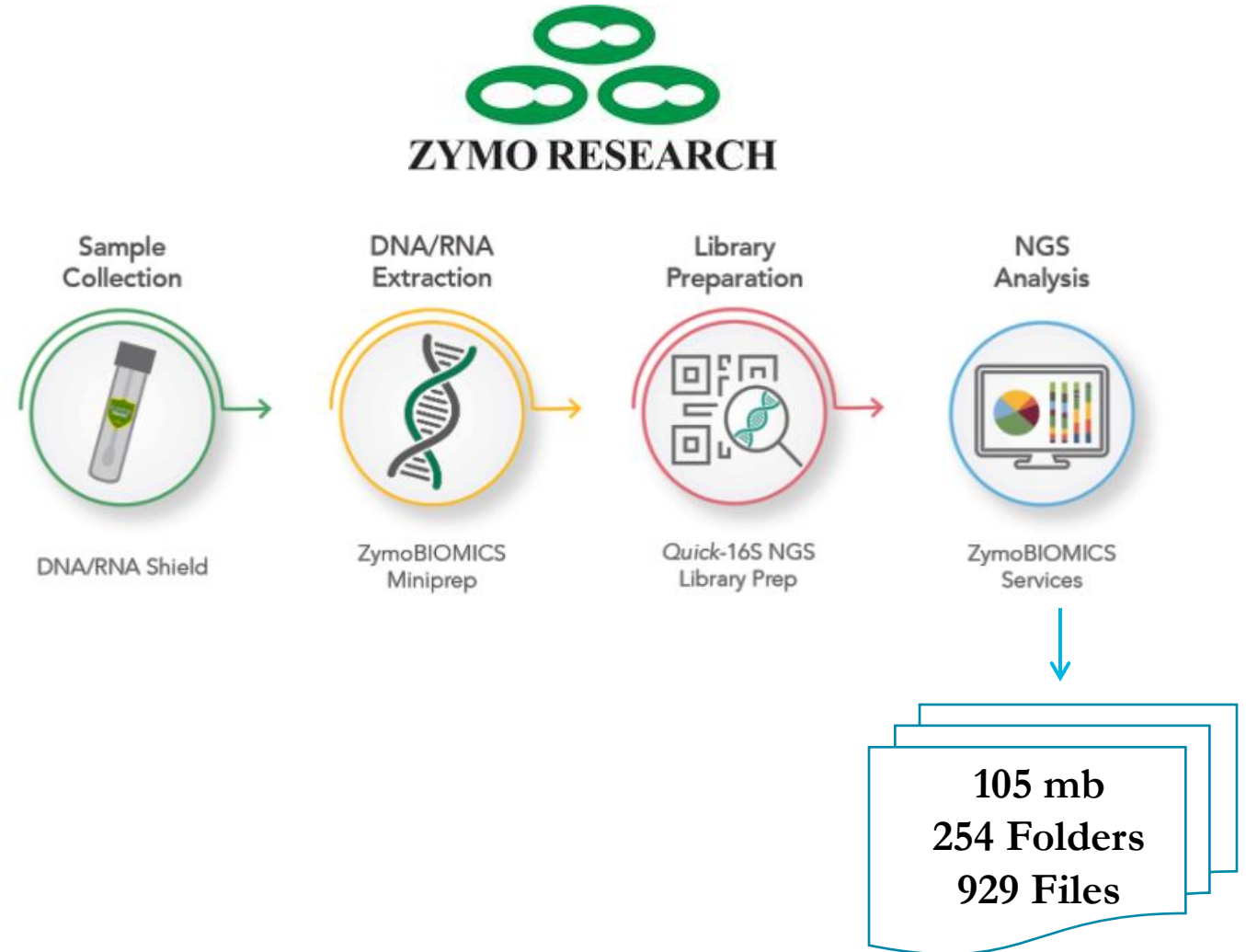
SWF Sampling Approach



- Entire column of SWF collected with pool skimmer net.
- SWF transferred into 2-gal Ziploc[®] bags.
- SWF refrigerated and allowed to condense for 24hrs.
- Slowly poured through cheese cloth into sample bottles to strain out large debris.
- 20mL condensed SWF preserved for Shotgun Metagenomic Sequencing.
- ≥ 20 mL condensed foam prepared for 41 PFAS analysis.

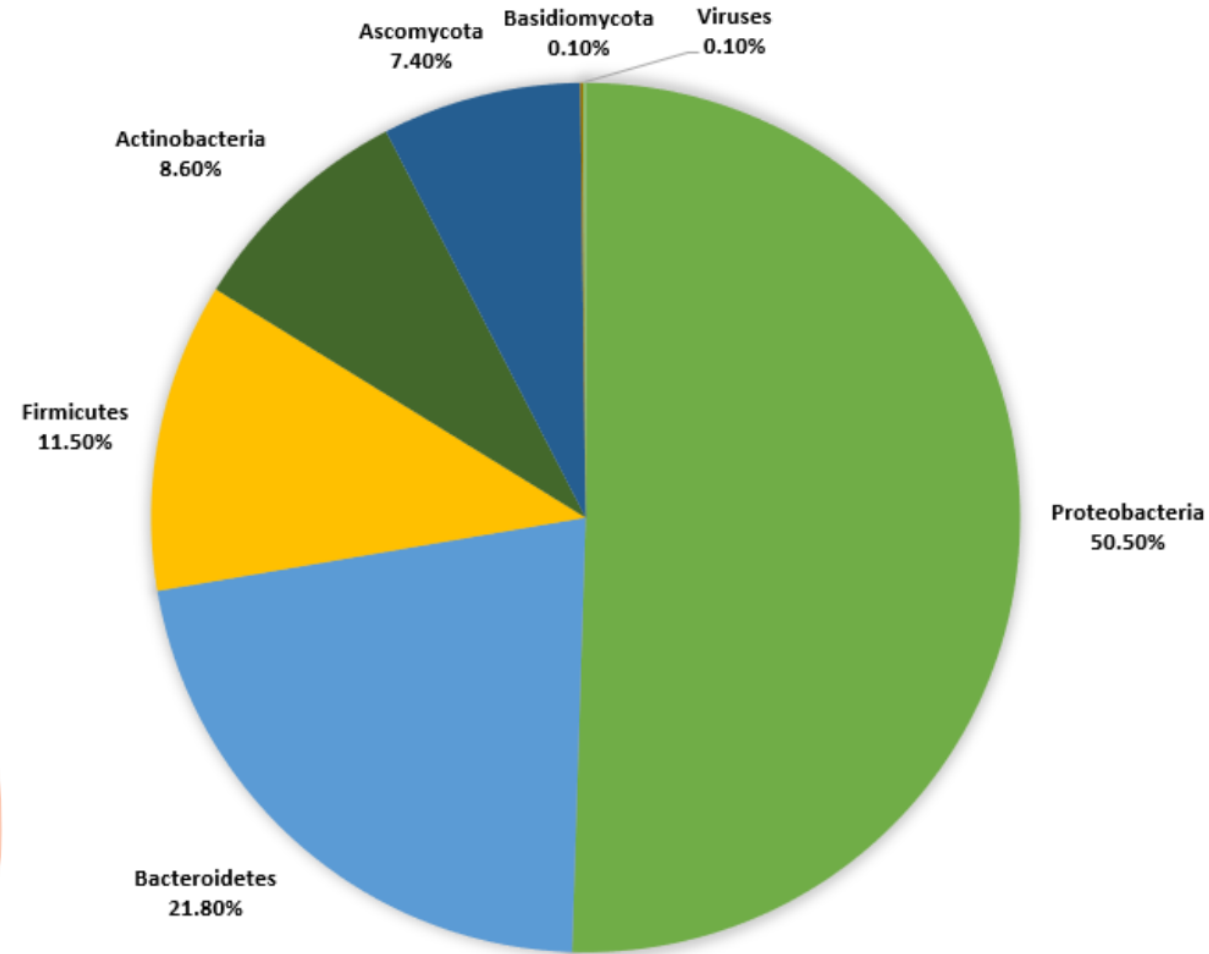
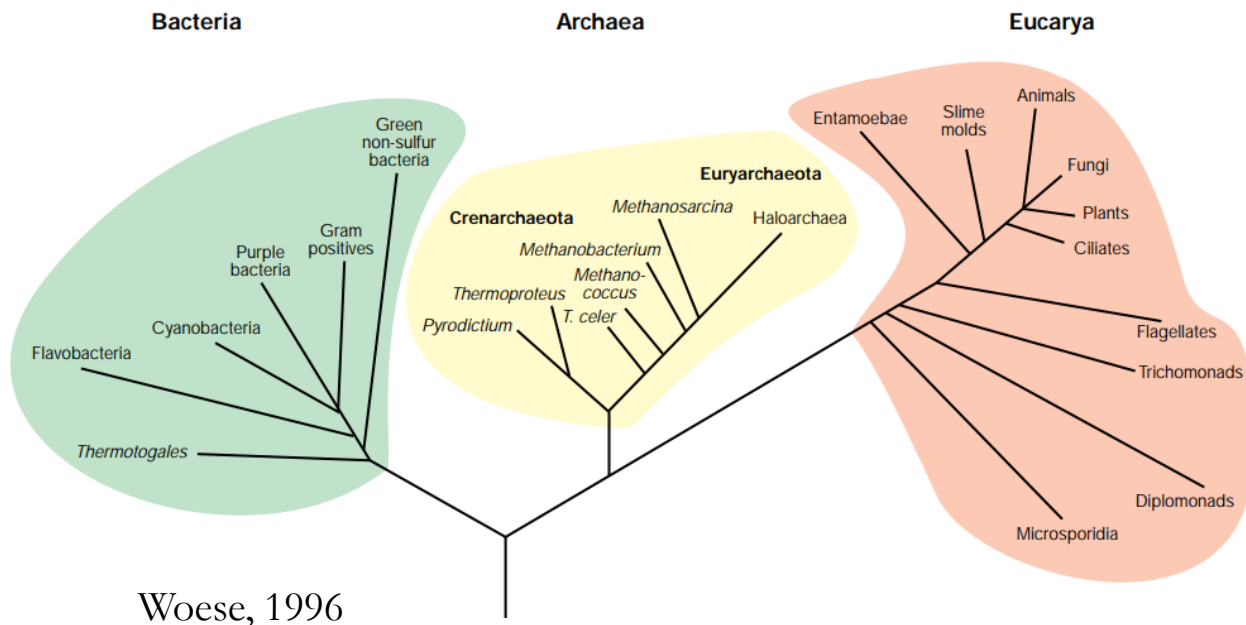
Shotgun Metagenomic Sequencing

- 15 SWF samples + 1 Duplicate
- ZymoBIOMICS®
Zymo Research, Irvine, CA
- Prokaryotes, fungi, limited virus
- Nextera® DNA Flex Kit
- Illumina NovaSeq®
- Trimmomatic Software Tool
DIAMOND Sequencer
QIIME Pipeline
DADA2



Predominant Phyla Identified in SWF

- Proteobacteria (50.5%)
- Bacteroidetes (21.8%); Firmicutes (11.5%)
- Actinobacteria (8.6%)
- Ascomycota – Fungi, including some yeasts



Phylogenetic Heat Map (Bacteria Strain Level)



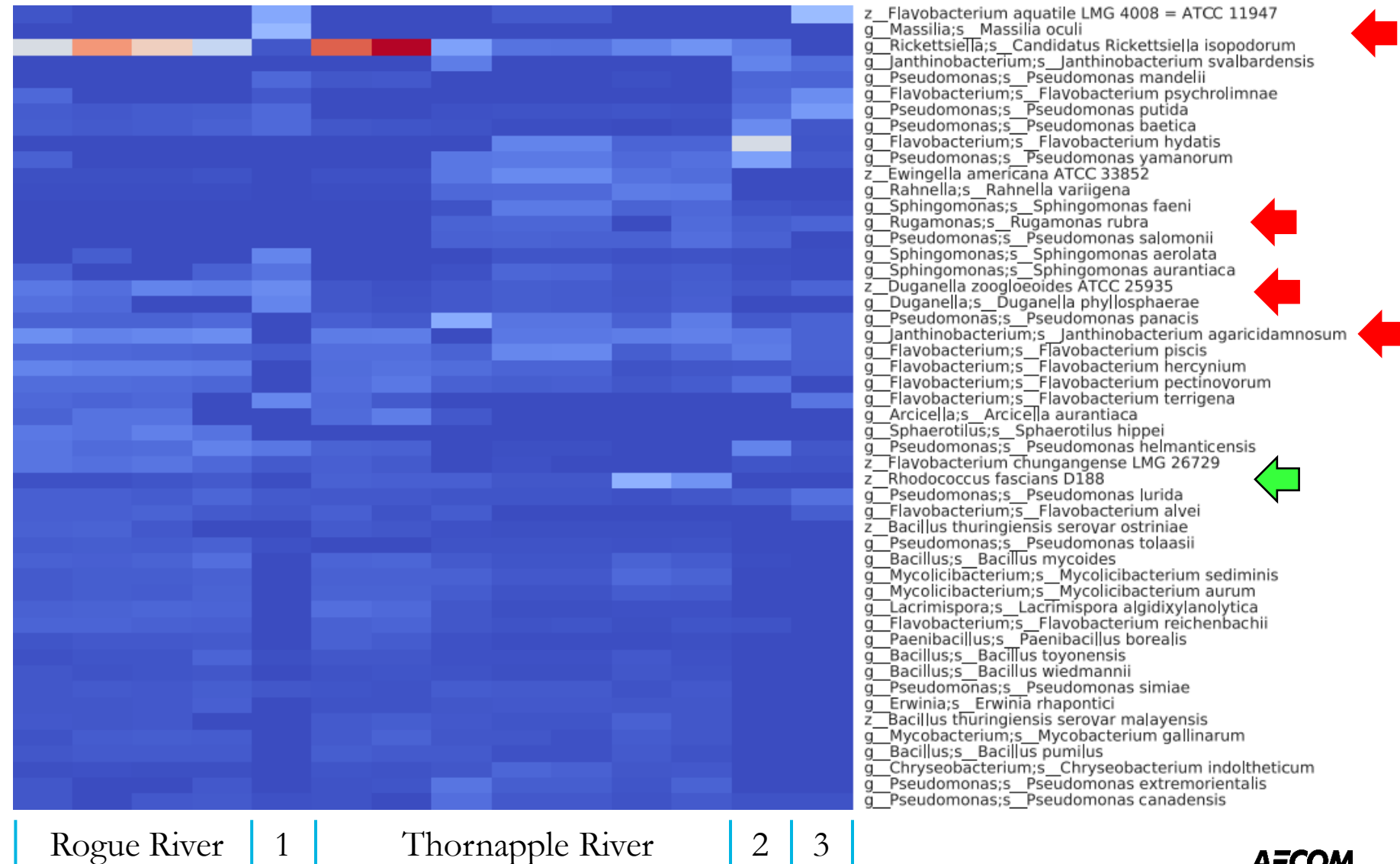
← “Oxalobacteria”

← “Rhodococcus”

1 – Huron River

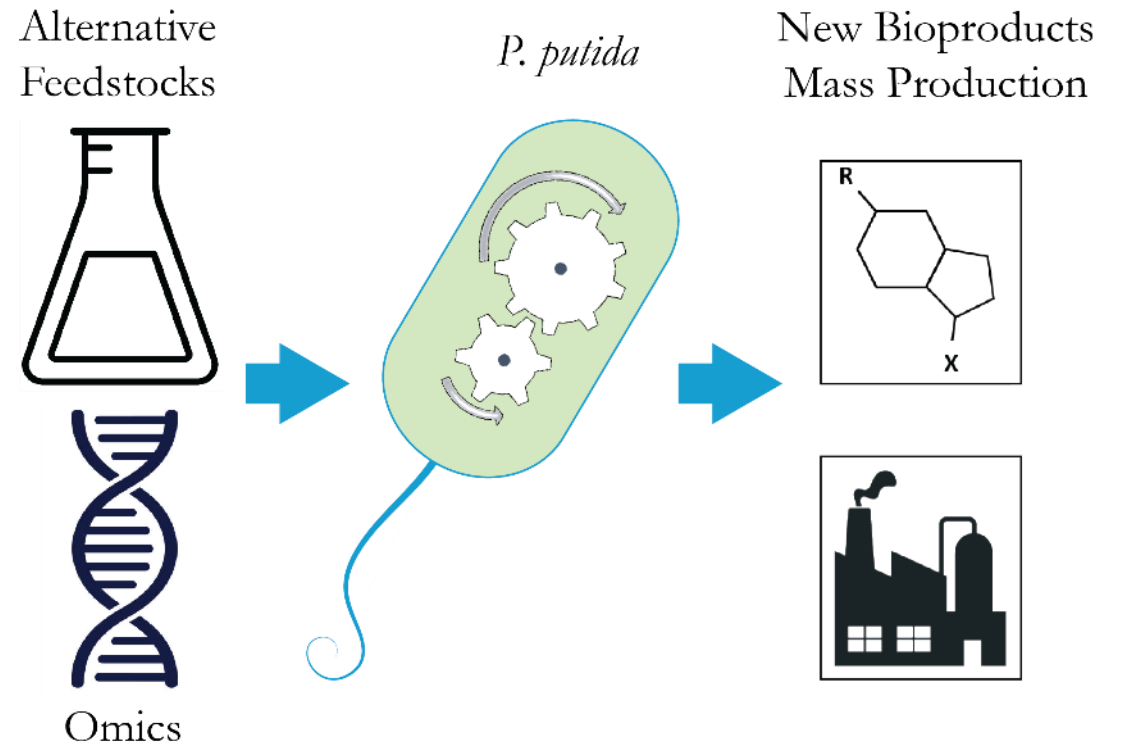
2 – Lake Margrethe

3 – Cedar Lake



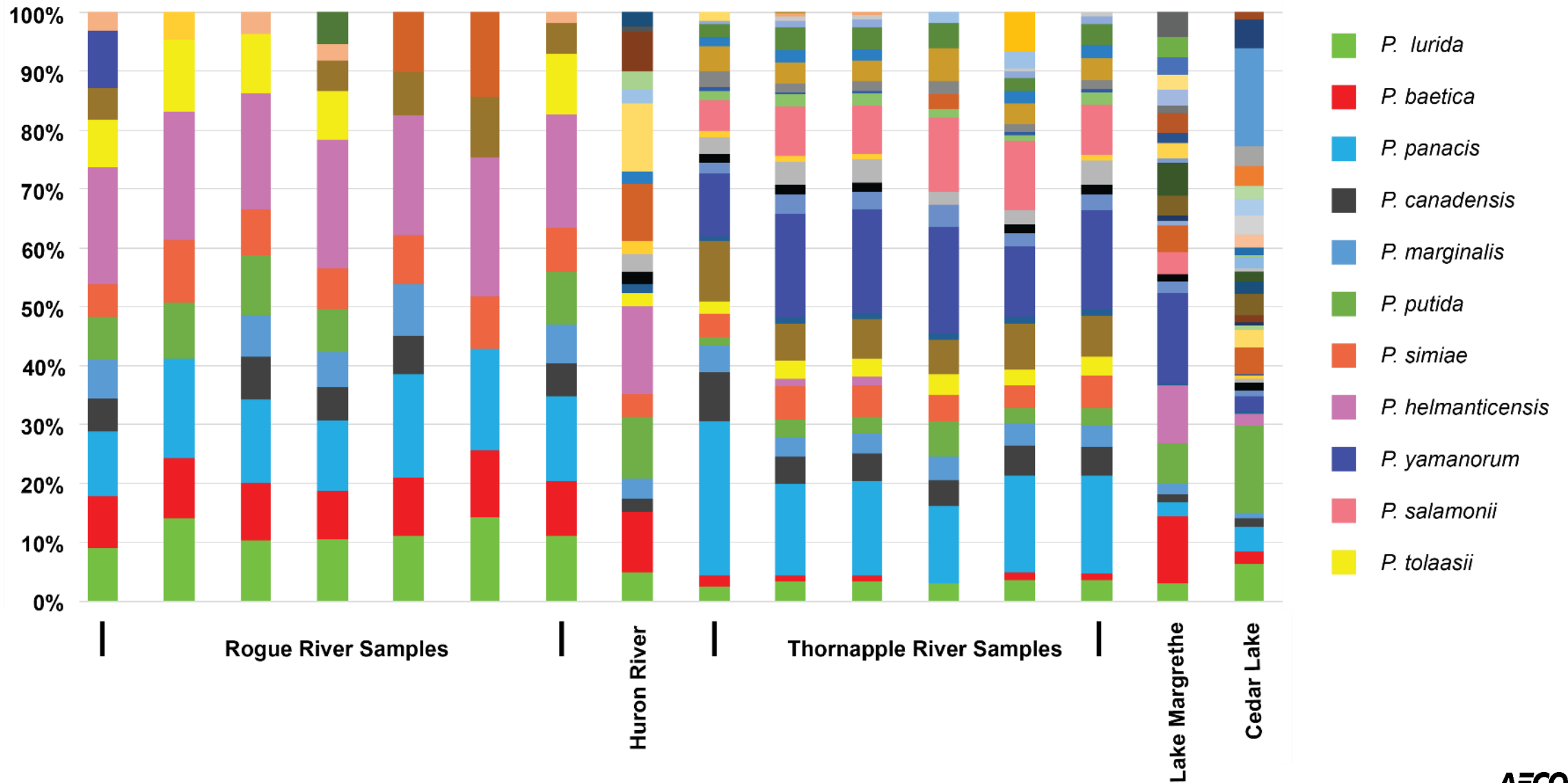
Pseudomonas sp.

- Common wide-spread genus of gram-negative gammaproteobacterial.
- Many documented as contaminant-dwellers and in bioremediation, including PFAS. Some produce biosurfactants.
- Have been identified in Feammox process (Sawayama, 2006).
- 191 validated species, some are now being lumped under *Burkholderia*.
- 63 species identified in SWF.
- *Pseudomonas lurida* found in all samples; *P. baetica*, *P. panicus*, *P. putida* and *P. helmanticensis* found in most of the samples.



Bacteria *P. Putida* as a functional chassis for creation of new biochemistries and products. Adapted from Nike & de Lorenzo, 2018.

Predominant *Pseudomonas* in SWF

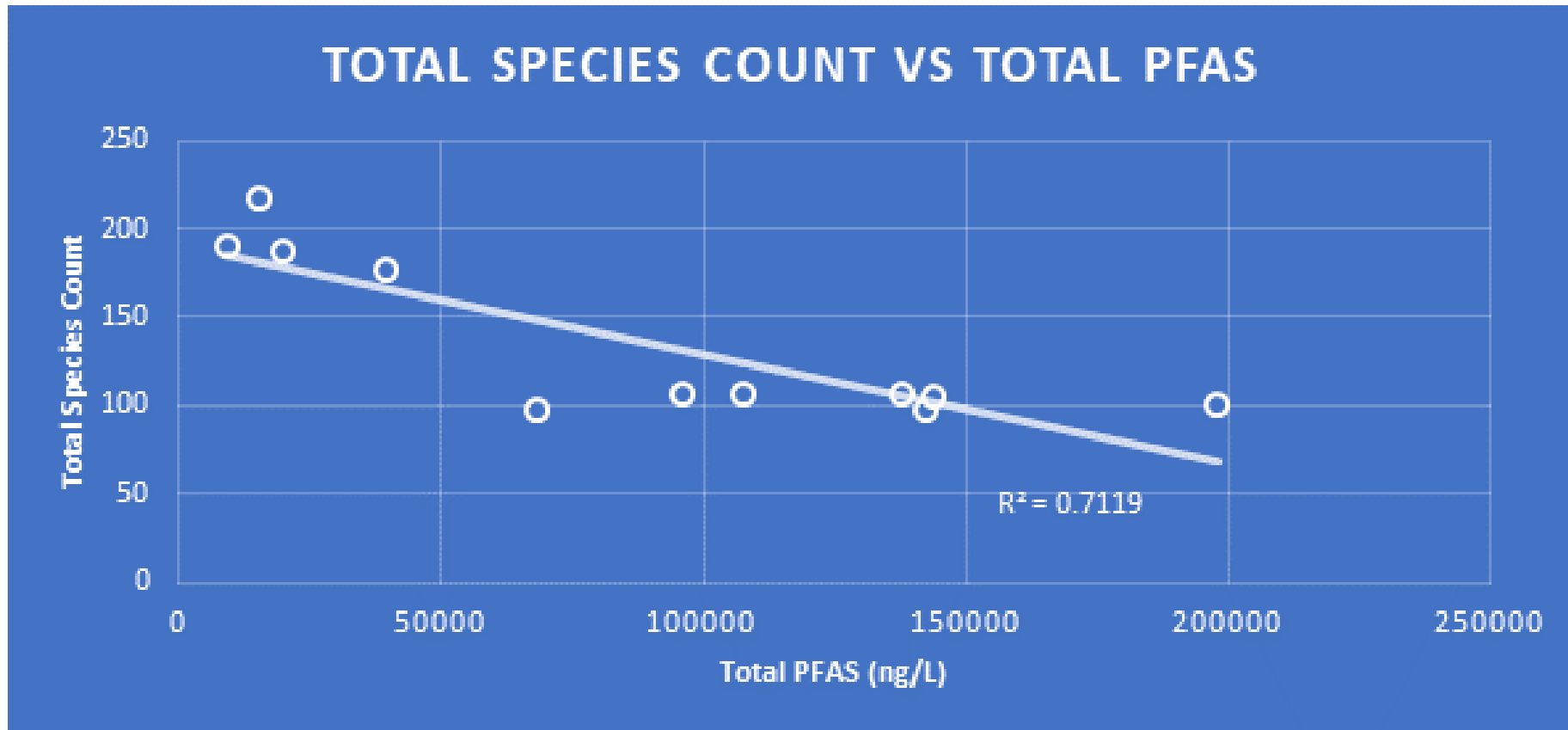


Functional Genes

- Over 750 functional genes identified in SWF, the most common genes summarized below.
- Dehalogenation genes not identified.
- Heme-b biosynthesis, involves use of **Uroporphyrinogen Decarboxylase (UroD)**.
 - Elimination of carboxyl groups; UroD performs decarboxylations without the intervention of any cofactors.
 - Considered the benchmark for catalytic efficiency in enzymes (Lewis & Wolfenden, 2008).

15 Samples	
Functional Gene / Species	Occurrence
HISDEG-PWY: L-histidine degradation I	11.0
PWY-6151: S-adenosyl-L-methionine cycle I	11.0
PWY-7111: pyruvate fermentation to isobutanol (engineered)	11.0
VALSYN-PWY: L-valine biosynthesis	11.0
HEMESYN2-PWY : heme biosynthesis II (anaerobic)	10.0
HOMOSER-METSYN-PWY: L-methionine biosynthesis I	10.0
PEPTIDOGLYCANSYN-PWY: peptidoglycan biosynthesis I (meso-diaminopimelate containing) g__Bacillus.s__Bacillus_cereus_thuringiensis	10.0
PWY0-1296: purine ribonucleosides degradation	10.0
PWY-6387: UDP-N-acetylmuramoyl-pentapeptide biosynthesis I (meso-diaminopimelate containing)	10.0
HEME-BIOSYNTHESIS-II : heme biosynthesis I (aerobic)	9.0

Total Number of Bacteria Species in River SWF
Plotted vs. Total PFAS Concentration (ng/L)



Q & A

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AECOM

THANK YOU

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